Received : 20 May 2024, Accepted: 15 June 2024 DOI: <u>https://doi.org/10.33282/rr.vx9i2.31</u>

GENETIC BLUEPRINTS AND BIOCHEMICAL MACHINERY: BIOTECHNOLOGY'S ROLE IN MODERN GENETICS

Rahmeen Ajaz¹, Maleeha Saleem², Tayyiba Mumtaz³, Laila Shawal^{4*}, Farina Kanwal⁵, Umber Rauf⁶, Sana Maryam⁷, Tahreem Arshad⁸, Nimra Ather⁹

¹Faculty of Public Health, Department of Occupational Health and Safety, Universitas Indonesia, Indonesia

²Department of Biochemistry, University of Agriculture Faisalabad

³Centre of Biotechnology and Microbiology, University of Peshawar

^{*4}Centre of Biotechnology and Microbiology, University of Peshawar

⁵Centre of Agricultural Biochemistry and Biotechnology, University of Agriculture Faisalabad

⁶Veterinary Research Institute, Zarar Shaheed Road Lahore Cantt, Punjab Pakistan

⁷Department of Pharmacology, The Islamia University Bahawalpur Pakistan

⁸Department of Pharmaceutics, The Islamia University Bahawalpur Pakistan

⁹Department of Zoology, Wildlife and Fisheries, University of Agriculture Faisalabad Pakistan

*Corresponding Author: Laila Shawal

Abstract

The field of genetics has undergone substantial shifts due to the combined activities of genetic programs and chemical processes, largely informed by advancements in biotechnology. This progression has evolved from simple genetic manipulation to groundbreaking innovations across various domains. Central to this evolution is the intricate interplay between the genetic maps conveyed by DNA sequences and the chemical dynamics governing gene regulation and function. Focusing on technological advancements with significant implications in biotechnology, notable developments include gene editing tools such as the CRISPR-Cas9 system, high-throughput DNA sequencing platforms, and RNA-based therapies. Recent advances have significantly improved our ability to handle and analyze genetic data. Biotechnology is crucial in areas like gene therapy and personalized medicine, which tailor treatments to an individual's genetic, refine gene-editing techniques, and lead to new medical, agricultural, and environmental innovations. The future potential of biotechnology in genetics is vast and promising.

Keywords: Biotechnology, Genetic Programs, Chemical Processes, Gene Editing, CRISPR-Cas9, Personalized Medicine



Graphical abstract

Introduction

The study of genetics has been scientifically revolutionized in the last few decades due to the advancement of biotechnology. The genetic map made up of DNA is the primary information base that builds the biochemical processes within a cell (Adlak et al., 2019). This has been an area of significant study; as a result, there have been major advancements and discoveries. In its defining aspect, genetics is a subdivision of biology that stems from the research of the Austrian geneticist Gregor Mendel, who conducted his work with pea plants in the 19th century (Gayon, 2016). However, the evolution of genetics started in the middle of the twentieth century when James Watson and Francis Crick presented the structure of DNA (Danylova & Komisarenko, 2020). This discovery revealed the helical structure of DNA and gave knowledge about how genetic information is stored and copied. Since then, genetics has grown immensely due to technological advancements that have enhanced more profound research into the molecular basis of heredity and variation (Moss, 2019).

ISSN : 2059-6588(Print) | ISSN 2059-6596(Online)

Genome is defined as the complete set of genetic instructions in an organism or the genetic makeup of an organism. It is made up of DNA, which is made up of long sequences of nucleotides in a particular order (Brown, 2023). Nucleotide comprises a sugar, a phosphate group, and a nitrogenous base. These bases include adenine (A), thymine (T), cytosine (C), and guanine (G), which form the sequence that determines the genetic information of the development and functioning of living organisms (Brown, 2020). The developments in sequencing and analyzing genomes have been one of the most significant leaps in genetics as it has given a complete look at the genetics of different organisms. Biotechnology in genetics can be illustrated by the Human Genome Project, accomplished in 2003 (Moraes & Góes, 2016).

This global research project sought to analyze and depict all the genes in the human body and their functionality (Ali et al., 2019). The end of the Human Genome Project signified the start of the post-genomic era when researchers started to study the genetic aspect of diseases, discover variations, and use genetics to provide customized treatment to patients. It also helped improve other fields of genomics, including comparative genomics, functional genomics, and epigenomics. Biotechnology has been central in making such developments possible (Stenson et al., 2017). The innovations in recombinant DNA technology in the 1970s enabled researchers to work on genetic material in previously impossible ways. This technology involves the integration of DNA from different sources to produce new sequences and allow the analysis and manipulation of genes (Lek et al., 2016). Molecular biology methods like gene cloning, PCR, and DNA sequencing have made working with specific genes and their products easier by amplifying, analyzing, or mutating particular DNA segments (Li et al., 2018).

The CRISPR-Cas9 system is one of the most groundbreaking biotechnological inventions in genetic engineering, as it is an efficient gene editing tool that can be utilized to modify the genome. Initially identified in bacteria and involved in the immune system of the bacteria, CRISPR-Cas9 has been employed in many organisms, including human beings (Tyagi et al., 2020). This technology consists of applying a guide RNA to take the Cas9 enzyme to a particular site on the genome, thus creating a double-stranded break (V. Singh et al., 2017). CRISPR-Cas9 has introduced new ways of experimentation in genetic research, ranging from genetically modified animal models for diseases affecting human beings to the generation of gene therapies for genetic diseases. Another significant advancement of biotechnology in contemporary genetics is the development of high-throughput sequencing technologies, or next-generation sequencing (NGS) (Hossain, 2021). Next-generation sequencing has allowed researchers to perform GWAS to find diseases' genetic markers and analyze gene expression and population differences (Nasykhova et al., 2019). The availability of large volumes of genetic data has also promoted the creation of bioinformatics tools and approaches to work with it. Biotechnology and genetic engineering have expanded the knowledge of the genetic basis of inheritance and implemented many applications in different fields. It has been applied in agriculture to produce genetically modified foods with favorable characteristics like resistance to pests, diseases,

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579 ISSN : 2059-6588(Print) | ISSN 2059-6596(Online) drought, and higher nutritional value (Black et al., 2015). These could increase yield, decrease reliance on chemicals, and solve problems of food insecurity when implemented (Alonso et al., 2015).

In medicine, biotechnology has improved the diagnosis and treatment of diseases. Diagnosis through genetic testing and screening has become the most efficient way of identifying patients at risk of inherited diseases so they can be treated early and efficiently (Splinter et al., 2018). Gene therapy, which involves replacing, modifying, or adding genes in a patient's body, holds promise for treating various genetic diseases. New techniques are more accurate, making treatments more effective. Personalized medicine, which tailors treatment based on an individual's genetic profile, is another important area of biotechnology and genetics. Both gene therapy and personalized medicine offer new ways to tackle diseases with greater precision." (Fernandez-Marmiesse et al., 2018).

Another area where biotechnology contributes a lot to genetics is in the synthetic biology discipline that merges engineering and biology paradigms. In this context, synthetic biology is defined as designing and constructing novel biological components, devices, and systems and modifying existing biological systems for specific uses (Goyal et al., 2022). With the help of this field, one can develop new biological products and processes that can dramatically change various sectors such as pharmaceutics, biofuels, etc. Ethical and safety issues are also observed in synthetic biology, which requires effective regulation of innovation (Madhavan & Mustafa, 2023). When exploring modern genetics technologies, it is essential to discuss the ethical, social, and consequential impacts of various biotechnology breakthroughs (Heams, 2015). Issues include the precision of controlling genetic material, potential side effects, genetic discrimination, and the use of human embryos. As we keep advancing in biotechnology, we must carefully consider these ethical and societal implications to ensure the benefits are used responsibly and fairly (Freemont & Kitney, 2015).

Genetic Blueprints: The Foundation of Life

Structure and Function of DNA

DNA, or deoxyribonucleic acid, can be defined as the hereditary material existing in almost all living organisms and being the basis of genetic information (Mattick & Amaral, 2023). The molecule's well-known double helix form, discovered by James Watson and Francis Crick in 1953, has two polynucleotide chains running in opposite directions and coiled around a central axis (Bretscher & Mitchison, 2017). These strands are made up of repeating units called nucleotides, each consisting of three components (Haidri et al., 2023): A phosphate group, a five-carbon sugar, deoxyribose, and one of four nitrogenous bases- adenine, thymine, cytosine, and guanine. The specifics of the two base pairings (for example, A-T and C-G) are that they create hydrogen bonds, which contribute to the stability of the DNA molecule. It permits the exact copying of genetic information (M. Tripathi et al., 2023).

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579

ISSN : 2059-6588(Print) | ISSN 2059-6596(Online)

The sequence of these nucleotide pairs contains the information required for all living organisms' growth, development, and functioning (Haidri, Ishfaq, et al., 2024). They are segments of DNA called genes, which are used in synthesizing RNA molecules with the help of transcription (Haidri, Qasim, et al., 2024). These RNA molecules, especially mRNA, play a role in transmitting genetic information from the DNA pool in the nucleus to the ribosomes in the cytoplasm for translation. Proteins are involved in many processes essential for the organism's life, they catalyze metabolic processes, replicate DNA, react to stimuli, and transport molecules (Javed et al., 2021). The study of DNA as hereditary material has been central to genetics and has helped explain how inherited information is stored and used (Ullah et al., 2024). The discovery of the double helix structure led to other scientific discoveries, such as molecular biology techniques like DNA sequencing, PCR, and genetic engineering (Hussain et al., 2024). These techniques have significantly transformed how scientists can analyze and edit genetic material, immensely impacting the medical, agriculture, and biotechnology fields (A. Singh et al., 2021).



Figure 1 Genetically modified organisms

The features of DNA sequencing have allowed individuals to discover the genes relevant to certain illnesses, comprehend the genetic variation among populations, and design treatment plans (Fatima et al., 2024). Biotechnology has produced genetically modified organisms (GMOs) with desirable characteristics like high yield and disease resistance (Riaz et al., 2023). DNA has also helped to reveal the history of evolution between different animals, giving molecular support to the theory of evolution. The current study of DNA formation and functioning proves

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579 ISSN : 2059-6588(Print) | ISSN 2059-6596(Online) that the study of genetics is a dynamic field that provides the basis for advancement in the life sciences (Riaz et al., 2021).

Gene Expression and Regulation

Gene expression is the information in a gene that is used to make a functional product (Hill et al., 2021). DNA carries genetic information, which is copied into messenger RNA (mRNA) using the enzyme RNA polymerase (Silverman et al., 2020). This involves opening the DNA double helix and using one strand to create an RNA molecule that matches the other strand. The mRNA sequence copies the gene's coding sequence and is used to make proteins. After it is made, the mRNA undergoes several changes, like capping the 5' end, splicing out introns, and adding a poly-A tail to the 3' end. These modifications stabilize the mRNA and help move it from the nucleus to the cytoplasm. In the cytoplasm, the ribosomes read the mRNA sequence and translate it into a specific sequence of amino acids to form a protein (Consortium, 2017).



Figure.2 From Gene to Protein: The Central Dogma of Molecular Biology

Protein synthesis involves transferring RNA (tRNA) molecules that bring the correct amino acids to the sequence encoded by the mRNA codons using the tRNA's anticodon sequence (Haidri, Fatima, et al., 2024). The ribosome helps link the amino acids by forming peptide bonds, creating a growing polypeptide chain that folds into a functional protein. Gene regulation controls when, where, and how much of a gene product is made. This regulation is essential for a cell's activities, growth, and response to its environment. There are different levels of regulation: transcriptional, post-transcriptional, translational, and post-translational (Ummer et al., 2023). At

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579

ISSN : 2059-6588(Print) | ISSN 2059-6596(Online)

the chromosomal level, transcription factors can attach to DNA sequences near genes, either helping or blocking RNA polymerase from starting transcription. Enhancers and silencers are DNA regions that can increase or decrease transcription levels when specific transcription factors bind to them (Gil & Ulitsky, 2020). Epigenetic modifications, like DNA methylation and histone modification, also regulate gene expression by changing chromatin structure and DNA accessibility. Post-transcriptional regulation happens after mRNA is created, involving processes like mRNA splicing, editing, and stability control. Alternative splicing allows different proteins to be produced from a single gene by including different exons in the final mRNA (Corbett, 2018). mRNA stability and degradation are controlled by RNA-binding proteins and microRNAs, which interact with mRNAs to affect their lifespan and translation rate (Feigerlovà & Battaglia-Hsu, 2017). Some regulatory elements located in the mRNA can influence the binding of the ribosomes and the start of the translation. Another level of regulation of protein function is post-translational modification of the proteins produced from the synthesized mRNA, which can include phosphorylation, ubiquitination, and glycosylation to change the activity, stability, location, and interactions with other molecules. Such changes enable cells to respond quickly to environmental changes by activating or inactivating specific proteins (Feigerlovà & Battaglia-Hsu, 2017). Gene regulation is a complex process that dictates where, when, and how much of any given gene is produced to create functional organisms. Knowledge of gene expression and regulation patterns aids in decoding the cell's functions and has vast applications in developmental biology, disease investigations, and biotechnology, among others (Feigerlovà & Battaglia-Hsu, 2017).

Biochemical Machinery: The Workhorses of the Cell

Enzymes and Proteins

Enzymes are proteins that act as catalysts, speeding up biochemical reactions without being used up. They are involved in many cellular processes like DNA synthesis, repair, transcription, and translation (Kornberg, 1991). Enzymes lower the activation energy needed for a reaction, thus increasing the reaction rate. They form enzyme-substrate complexes by binding to the substrate at the active site, helping to convert substrates into products through methods like induced fit and transition state stabilization (Shanmugam, 2009). For example, during DNA replication, enzymes like DNA polymerase add nucleotides to create new DNA strands using a template strand. Helicase separates the DNA strands, primase makes RNA primers to start replication, and ligase joins breaks in the DNA to ensure continuity (Anita et al., 2024). In DNA repair, some proteins include DNA ligase, endonucleases, and exonucleases, which are responsible for identifying the errors or damage in the DNA and repairing them to ensure that the genetic material is not compromised. RNA polymerase catalyzes to convert RNA from a DNA template during transcription (Fossel, 2015). There are also proteins known as transcription factors that help initiate and control transcription so that the genes to be expressed are expressed at the right time. Likewise, enzymes such as ribonuclease and spliceosome complexes contribute to the maturation of RNA molecules in preparation for translation to protein products. Proteins, the gene products, assume various functions within the cell, as bounded by their structure and characteristics. Connective tissue proteins like collagen and keratins help support and rigidify the tissue and cells to give strength and structure (Sindhu et al., 2022).

Disease/Condition	Cause/Pathology	Therapeutic	Ref.	
		Enzymes [Brand]		
α-Mannosidosis	Deficiency of α -D-	Velmanase α	Mehta & Beck (2020)	
	mannosidase	[Lamzede]		
Batten disease	Deficiency of	Cerliponase a	Schulz et al. (2013)	
	tripeptidyl peptidase	[Brineura]		
	1			
Pompe's disease	Deficiency of acid α-	α-glucosidase	Kishnani & Howell	
	glucosidase	[Myozyme]	(2004)	
Metabolic				
Deficiencies				
Exocrine pancreatic	Insufficient	Pancreatic enzymes	Borowitz & Stevens	
insufficiency (EPI)	pancreatic enzymes	[Enzepi]	(2012)	
Phenylketonuria	Deficiency of	PAH and	Blau & van Spronsen	
(PKU)	phenylalanine	phenylalanine	(2010)	
	hydroxylase (PAH)	ammonia-lyase		
		[Palynziq]		
Severe combined	Deficiency of	Polyethylene glycol-	Hershfield (1995)	
immunodeficiency	adenosine deaminase	conjugated ADA		
(SCID)	(ADA)			
Wolman disease	Deficiency of	Lysosomal acid	Pisciotta & Busnelli	
	lysosomal acid lipase	lipase [Kanuma]	(2017)	
Acute intermittent	Deficiency of	Hydroxymethylbilane	Anderson & Sassa	
porphyria (AIP)	hydroxymethylbilane	synthase and	(2006)	
	synthase	porphobilinogen		
		deaminase		
Congenital sucrase-	Deficiency of sucrase	Sacrosidase	Treem (1995)	
isomaltase deficiency	and isomaltase			
(CSID)				
Hypophosphatasia	Deficiency of	TNSALP [Strensiq]	Whyte & Greenberg	
	alkaline phosphatase		(2012)	
	(TNSALP)			
Protein C deficiency	Deficiency of Protein	Protein C [Ceprotin]	Griffin & Evatt	

 Table.1 Therapeutic Enzyme Applications for Various Diseases and Conditions

Remittances Review June 2024, June: 9. No: 3, pp.539-579

remittancesreview.com

Volume:	9, No: 3,	pp.539-579
ISSN : 2059-6588(Print) ISS	N 2059-6	596(Online)

	С		(1992)
Lactose intolerance	Reduction of lactase	Lactase	Woteki & Thomas
	activity		(1998)
Fibrosis Conditions			
Chronic total	Fibrous plaques in	Collagenase	de Oliveira &
occlusions	coronary arteries	Clostridium	Silveira (2014)
		histolyticum (CCH)	
Dupuytren's disease	Thickening of fascia	Collagenase	Badalamente & Hurst
	tissue in hands	Clostridium	(2007)
		histolyticum (CCH)	
		[Xiapex]	
Peyronie's disease	Fibrous plaques in	Collagenase	Gelbard & Jarow
	the penis	Clostridium	(2007)
		histolyticum (CCH)	
Uterine fibroid	Fibroid tissue growth	Collagenase	Ulbrich & von
	around the uterus	Clostridium	Rappard (2010)
		histolyticum (CCH)	
Keloid disease	Overgrowth of scar	Collagenases and	Bloemen & van der
	tissue	matrix	Wal (2009)
		metallopeptidases	
Lung cystic fibrosis	Viscous secretions in	Deoxyribonuclease I	Yankaskas &
	lungs	[Pulmozyme]	Marshall (2004)
Glaucoma	Fibrous formations at	Collagenases	Coleman & Migdal
	trabecular meshwork		(2008)
Ocular Affections			
Various ocular	Malfunction of	Chondroitinase,	Avery (2009)
diseases	vitreous humor	hyaluronidase,	
		nattokinase,	
		ocriplasmin [Jetrea]	

The cytoskeleton, composed of actin and tubulin, is essential for cell movement, division, and transport (Aseervatham, 2020). Proteins like hormones and receptors help cells and tissues communicate and change their functions. For example, insulin helps glucose enter cells, and neurotransmitter receptors manage signals between neurons. Signaling pathways involve interactions between proteins and phosphorylation, where kinases add phosphate groups to regulate actions (Biswas et al., 2022). Metabolic pathways are vital for cell survival, and enzymes drive these processes by breaking down nutrients like carbohydrates, fats, and proteins for energy and other uses. Amylase breaks down starch into sugars, lipase breaks down fats into

fatty acids and glycerol, and proteases break down proteins into amino acids (Pizzagalli et al., 2021). Transport proteins, such as hemoglobin and membrane transporters, are crucial for moving molecules and ions within and between cells. Hemoglobin carries oxygen from the lungs to tissues, while membrane transporters move nutrients into cells and waste out (Cherian, 2022). Proteins also play a role in the immune response, with antibodies identifying and neutralizing pathogens. Regulatory proteins control gene expression, cell division, and cell death (Nair, 2016).

Proteins have diverse functions due to their structures, which are determined by genes and amino acid sequences. They form specific shapes through interactions like hydrogen bonds and ionic bonds. Changes or mutations in proteins can lead to diseases like cystic fibrosis, Alzheimer's, and cancer. Understanding enzymes and proteins is crucial for understanding cell processes and molecular organization. This knowledge has significantly advanced medicine, biotechnology, and bioengineering, leading to enzyme therapy, drug design, and improved biomaterials (Maiuri & Kroemer, 2018).

RNA and Ribosomes

RNA molecules are very crucial in the transfer of genetic information from DNA to proteins. Different RNA molecules include mRNA, tRNA, rRNA, and micro-RNA, all involved in gene expression. mRNA transports the copied code from DNA to those structures called ribosomes, where the code is manifested into proteins. tRNA transports suitable amino acids to the ribosome during translation and ensures that the appropriate amino acids are attached in sequence to the polypeptide chain, depending on the template mRNA (Catalanotto et al., 2016). The most well-known rRNA is the molecular part of ribosomes. In contrast, these particles are unique complexes that synthesize new proteins according to the information in a message, an mRNA molecule (Scherrer, 2018).

Ribosomes are large structures that are made up of rRNA and proteins. These are two subunits, the large and the small, and they combine during translation. The ribosome brings mRNA and tRNA molecules together so that the process of mRNA translation into a polypeptide chain can occur. The small subunit interacts with the mRNA, and the large subunit contributes to forming peptide linkages between the amino acids (de Farias & José, 2020). This process involves three main steps: This has been divided into three phases: initiation phase, elongation phase, and termination phase. In initiation, the ribosome comes around the target mRNA, and the first tRNA is attached to the start codon. When the process is in the elaboration phase, the ribosome translates along the mRNA to codons where the appropriate aminoacyl-tRNA will form the new polypeptide chain. In termination, the ribosome encounters a stop codon, releasing the newly synthesized protein while the ribosome falls apart (Shen et al., 2018).



Figure3 Bacterial mechanism in the antibiotics process (Baran et al., 2023)

Recombinant DNA Technology

Recombinant DNA technology or rDNA technology uses laboratory techniques to combine and restructure DNA in an organism, thus even modifying an organism's genetic material. This field has evolved genetic research and technological applications that offer instruments for scientists to analyze and manage genetic operations as they choose. The main ideas of recombinant DNA technology are gene cloning, PCR, polymerase chain reaction, and CRISPR-Cas9 (Sivamani et al., 2024). Gene cloning is a process in which a gene of interest is placed into a vector, most commonly in the form of a plasmid, and then introduced into the host organism, often bacteria. This makes it possible to isolate and magnify the particular gene of interest to help investigate processes it controls and regulates (T. A. Singh et al., 2020). Cloning is helpful as it provides the researcher with large quantities of a gene or the protein product of the gene for use in gene therapy, the production of pharmaceuticals, and agriculture. PCR is one of the other fundamental methods that facilitate amplifying specific DNA sequences, starting with a small amount of the source material. This involves heating and cooling cycles where the DNA strands are denatured, annealed, and extended, and as such cycles are repeated, there is exponential replication of DNA. PCR has been established as one of the central technologies in molecular biology used to amplify DNA in diagnostics, forensics, and evolution studies (Bhatia & Goli, 2018).



Fig 4. Process involved in recombinant DNA technology

CRISPR-Cas9 is another widely innovative development in recombinant DNA technology that came later. A gene-editing tool, CRISPR-Cas9, is a bacterial immune system that can efficiently target and cut DNA sequences (Tüzmen et al., 2018). A guide RNA recognizes the target DNA sequence and can guide the Cas9 endonuclease to specific loci within the genome to create double-strand breaks. This allows for particular manipulations, for example, gene knockouts, insertions, or corrections, which has transformed the analysis of gene function and the creation of gene therapies for genetic diseases. The application aspect is not limited to research; medicine, agriculture, and industry are some areas that have been affected by recombinant DNA technology (Puvanakrishnan et al., 2019).

				r					
Organism Main		Model	Desci	ription	Refe	renc	e		
		Products	/Applications						
E. coli	K-12	Biofuel,	multipurpose	iML1515	1515	genes,	Lee	&	Kim
MG1655		recombin	ant proteins		2719	reactions,	2020		
					1192				
					metał	olites			

Table.2 Genome	e-Scale Metabol [®]	ic Models of	[•] Industrial P	latform Strains
Tuble Contine			Industrial I	

Remittances Review

June 2024

			June 2024,
		Volume: 9,	No: 3, pp.539-579
	ISSN : 2059	-6588(Print) ISSN 2	2059-6596(Online)
Alcoholic beverages,	Yeast8,	1133 genes,	Sandberg et al.,
bakery products,	ecYeast8,	3949 reactions,	2019
bioethanol	panYeast8,	2680	
	coreYeast8,	metabolites	
	proYeast8		
Amino acids	iCW773	773 genes,	Ohno et al.,
		1207 reactions,	2017
		950 metabolites	
Industrial enzymes and	iBsu1144	1144 genes,	Pfeifer et al.,
antibiotics		1955 reactions,	2017
		1103	
		metabolites	
Resveratrol	iYL1539	1539 genes,	McCloskey et
		2255 reactions,	al., 2018
		2231	
		metabolites	
Antibiotics, secondary	Sco-GEM,	1777 genes,	Ajikumar et al.,
metabolites	EcSco-GEM	2612 reactions,	2010
		2073	
		metabolites	
Lipids, pigments for	iCZ946	946 genes,	Kumar &
biofuel, food		2294 reactions,	Singh, 2017
supplements		1770	
		metabolites	

C alutamicum	Amino acida	iCW773	773 genes	Ohno et al
C. giutainicuin	Ammo actus	IC VV //3	1207 resistions	01110 et al.,
AICC15052			1207 reactions,	2017
			950 metabolites	
B. subtilis	Industrial enzymes and	iBsu1144	1144 genes,	Pfeifer et al.,
	antibiotics		1955 reactions,	2017
			1103	
			metabolites	
Alternaria sp.	Resveratrol	iYL1539	1539 genes,	McCloskey et
MG1			2255 reactions,	al., 2018
			2231	
			metabolites	
S. coelicolor	Antibiotics, secondary	Sco-GEM,	1777 genes,	Ajikumar et al.,
	metabolites	EcSco-GEM	2612 reactions,	2010
			2073	
			metabolites	
C. vulgaris	Lipids, pigments for	iCZ946	946 genes,	Kumar &
	biofuel, food		2294 reactions,	Singh, 2017
	supplements		1770	_
			metabolites	
L.	Starter in food	iLM.c559	559 genes,	Lee et al., 2011
mesenteroides	fermentation (dairy,		1088 reactions,	
subsp. cremoris	meat, vegetable		1129	
ATCC 19254	products)		metabolites	
L. reuteri JCM	Starter in food	Lreuteri_530	530 genes, 710	Michalak &
1112	fermentation, probiotic		reactions, 658	Brigham, 2015
	products, reuterin		metabolites	-
N. salina	Lipids, pigments for	iNS934	934 genes,	Lee et al., 2018
	biofuel, food		2345 reactions,	
	supplements		1985	
			metabolites	
C. reinhardtii	Biofuel	nd*	3726 reactions,	Sandberg et al.,
			2436	2020
			metabolites	
			1	

S. cerevisiae

In medicine, this technology has led to new treatments, such as recombinant insulin and other therapeutic proteins, and advancements in gene therapy to fix genetic disorders. In agriculture, scientists have created genetically modified crops with better traits, like resistance to pests, diseases, and drought, as well as improved nutritional value. These crops help feed the world and support sustainable farming (Arjmand et al., 2020). Recombinant DNA technology also allows us to make biofuels, bioplastics, and other products using specially engineered microorganisms. As the field develops, we expect to discover more methods and applications that will address global challenges and improve people's lives (Bhoria et al., 2022).

Gene Editing and CRISPR-Cas9

CRISPR-Cas9 is a powerful and precise tool for editing genes that has transformed genetic research and biotechnology. CRISPR stands for Clustered Regularly Interspaced Short Palindromic Repeats, and Cas9 is the enzyme that acts like a molecular scissor guided by RNA (Shah et al., 2018). The guide RNA finds a specific DNA sequence in the genome, and then the Cas9 enzyme cuts that exact spot. This cut allows scientists to add, delete, or replace genetic material, enabling targeted changes to the genome. CRISPR-Cas9 has many uses (Akram et al., 2023). It helps researchers study gene functions more effectively by creating specific gene knockouts or mutations. This has increased our understanding of gene regulation, development, and genetic diseases. Additionally, CRISPR-Cas9 is used in functional genomics to perform high-throughput screening, helping to identify genes that control various cellular processes and diseases (Wang et al., 2018).

Disease/Disorde	Model/Syste	Target	Gene	Editing Method	Reference
r	m	Gene(s)	Function		S
Rheumatoid	RAW264	miR-155	Pro-	Knockout	Jing et al.,
Arthritis (RA)	Cells		inflammatory		2015
			regulation in		
			RA		
Hereditary	Mouse	FAH	Tyrosine	Gene editing	Yin et al.,
Tyrosinemia			catabolism		2016
Type 1 (HT-1)			pathway,		
			toxic		
			accumulation		
Mice	Hpd	Gene	Pankowicz et		
		disruption	al., 2016		
Ornithine	Mice	OTC	Urea cycle	Gene editing	Yang et
Transcarbamylas			involvement		al., 2016
e (OTC)					

Fable 3 Gene Editing ir	Various Disease N	Iodels: Target G	enes, Functions,	and Methods
--------------------------------	-------------------	------------------	------------------	-------------

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579

ISSN : 2059-6588(Print) | ISSN 2059-6596(Online)

Deficiency					
Arginase	iPSC Cells	Arg1	Final step in	Gene knock-in	Lee et al.,
Deficiency			urea cycle		2016a
			control		
Duchenne	Mouse	Dmd	Dystrophin	Gene	Zhu et al.,
Muscular			protein	addition/deletion	2017; Li et
Dystrophy			truncation		al., 2015a
(DMD)					
Skeletal Muscle				El Refaey et al.,	
Stem Cells				2017	
Limb Girdle	Mice, iPSC	DYSF, α-	Dysferlin	Gene correction	Turan et
Muscular	Stem Cells	sarcoglyca	production		al., 2016
Dystrophy Type		n	stabilises		
2B (LGMD2B)			dystrophin		
Diabetes Mellitus	Mouse	DMPK	Disease	Gene editing	van
Type 1 (DM1)			aetiology		Agtmaal et
					al., 2017
Huntington's	Fibroblasts,	HTT	Brain	Gene editing,	Monteys
Disease (HD)	Mouse		function	ORF deletion	et al.,
			regulation;		2017;
			huntingtin		Talan,
			synthesis		2015
Friedreich Ataxia	Mouse	Frataxin	Mitochondria	Gene editing	Ouellet et
(FA)			l oxidative		al., 2017
			stress		
			involvement		
Amyotrophic	iPSC Cells	SOD1,	RNA/DNA	Gene editing	Wang et
Lateral Sclerosis		FUS	binding		al., 2017
(ALS)			proteins		
Cystic Fibrosis	iPSC Cells	CFTR	Chlorine	Base editing	Firth et al.,
(CF)			transport		2015
			regulation		
Alpha-1	iPSC Cells	AAT	Serum	Gene disruption	Smith et
Antitrypsin			trypsin	-	al., 2015
Deficiency			inhibitor		
(AATD)					
Recessive	Mice	COL7A1	Collagen	Gene disruption	Hainzl et
Dystrophic			production	*	al., 2017
Epidermolysis			_		

Remittances Review

June 2024, Volume: 9. No: 3. pp.539-579

	// [-]
ISSN : 2059-6588(Print) ISSN	2059-6596(Online)

Bullosa (RDEB)					
Dominant	iPSC Cells	COL7A1	Collagen	Mutant allele	Shinkuma
Dystrophic			production	disruption	et al.,
Epidermolysis					2016
Bullosa (DDEB)					
Multidrug	Escherichia	ft, and,	Cell division	Gene	Gomaa et
Resistance in	coli, Galleria	msbA,	control,	knockout/deletio	al., 2014;
Infectious	mellonella	nusB, ease,	bacterial	n, disruption	Citorik et
Diseases		blaSHV-	population,		al., 2014;
		18,	drug		Yosef et
		blaNDM1,	resistance		al., 2015;
		gyrA,			Kim et al.
		NDM-1,			
		CTX-M-			
		15,			
		blaTEM,			
		blaSHV,			
		aph-3,			
		mecA			
Acquired	CHME5	LTR U3	Viral	Knockout	Hu et al.,
Immunodeficienc	Cells, Mice	region,	transcription		2014; Yin
y Syndrome		gag, pol	al activity		et al.,
(AIDS)			control		2017a;
					Bella et
					al., 2018
Burkitt's	SNU-719	BART5,	Capsid	Gene deletion	van
Lymphoma,		BART6	protein		Diemen et
Hodgkin's			expression		al., 2016
Disease			control		
Herpes Simplex	TC620 Cells	ICP0,	Capsid	Indels	Roehm et
Virus (HSV)		ICP4,	protein		al., 2016
Infection		ICP27	expression		
			control		

In therapeutic approaches, the potential of CRISPR-Cas9 can be described as enormous in treating genetic diseases. With this technology, it is possible to treat diseases such as cystic fibrosis, sickle cell anemia, and muscular dystrophy, permanently since the mutations are

corrected at the DNA level (Abdelnour et al., 2021). Clinical trials are currently being conducted for CRISPR-based treatments, and the initial findings are quite encouraging. Also, the use of CRISPR-Cas9 in agriculture, for instance, is to modify crops to have better yield, disease-resistant crops, and crops that can be resistant to drought (Bhattacharjee et al., 2022). It is also used in synthetic biology and designing genetically modified organisms to manufacture biofuels, drugs, and other valuable bioproducts. CRISPR-Cas9 is easy to use, can target any gene, and has been made available globally for laboratories, boosting innovation in many areas. As researchers improve this method and sort out ethical and safety issues, CRISPR-Cas9 is set to be one of the leading technologies in genetic engineering that can bring changes to medicine, agriculture, and biotechnology that are expected to transform the world (Luthra et al., 2021).

Genomics and Bioinformatics

High sequencing technologies have transformed genomics, the study of entire genomes. These technologies allow scientists to sequence and compare large amounts of genetic data quickly and affordably (R. Tripathi et al., 2016). Next-generation sequencing (NGS) tools help scientists sequence entire genomes from humans to microorganisms, enhancing our understanding of genetic structures, functions, and variations (Raza & Ahmad, 2019). Genomics has several areas, including whole-genome sequencing, genome-wide association studies (GWAS), and comparative genomics. Genome sequencing helps identify genetic differences such as SNPs, insertions, deletions, and structural variations that are linked to diseases (Nazipova et al., 2018). GWAS uses extensive genomic data to find connections between genetic markers and traits, improving our knowledge of genetic risks and supporting personalized medicine (Pereira et al., 2020).

Bioinformatics is a fast-growing field in biology and medicine that involves organizing and analyzing genomic data. It uses computational tools and methods to analyze and visualize data, predict gene functions, and study species' evolutionary relationships. Bioinformatics applications include sequence identification, gene expression analysis, protein folding, and gene function prediction. Tools and databases like BLAST, Ensembl, and UCSC Genome Browser are commonly used for these purposes (Javed et al., 2021). Bioinformatics also integrates genomics with other fields like transcriptomics, proteomics, and metabolomics, providing deeper insights into biological processes and disease mechanisms (Ari & Arikan, 2016).

Applications of Biotechnology in Modern Genetics

Genetic Engineering and GMOs

Biotechnology has enhanced the field of genetic engineering, which creates new GMOs with various favorable characteristics to solve diverse issues in agriculture and food production. Genetic engineering is a method of direct intervention in the organism's genome through technology to insert, augment, or alter particular genes. This process generally involves

techniques like recombinant DNA technology, gene editing, and transformation to induce the intended genetic changes (Abdul Aziz et al., 2022). Genetic engineering is widely used to produce genetically modified crops, also called bio crops. Techniques like genetic engineering allow crops with specific characteristics, including pest resistance, herbicide tolerance, and enhanced nutritional qualities (Javed et al., 2021). For example, Bt cotton and Bt corn are genetically modified to produce proteins from the bacterium *Bacillus thuringiensis*, which is deadly to certain insects but harmless to man and animals. This trait helps cut the use of chemical pesticides, benefiting the environment and the wallet. Likewise, crop plants with herbicide tolerance allow farmers to use herbicides to control weeds, increasing the yield and decreasing the cost of agriculture (Yali, 2022).



Figure 5. Genetically modified crops are more resilient to environmental impacts

Another improvement is that foods' nutritional value has been improved. Some transgenic crops like Golden Rice have been bioengineered to contain higher provitamin A, or beta-carotene, to help eliminate provitamin A deficiency in developing countries. Such modifications can significantly influence public health since the quality of staple foods eaten by a vast population is enhanced. Some of the effects of genetic engineering and GMOs are not limited to agriculture only (Javed et al., 2021). Besides improving the ability of crops to withstand biotic and abiotic stresses and enhancing their nutritional value, such technologies can be helpful in the management of the physical environment and sustainable practices (Pereira et al., 2020). For instance, biotechnological applications such as genetically modified microorganisms can be used

in bioremediation to help remove substances that pollute the environment. These microorganisms can break down pollutants better than other microorganisms. Moreover, it is applied in synthesizing drugs and industrial enzymes, enhancing the production of medicine and biotechnology (Napier et al., 2019).

Table4.	Comprehensive Overview	of Gene Editi	ng Techniques	s for Disea	se Resistance	and
Nutritio	nal Enhancement in Agricul	lture and Live	estock			

Trait Category	Target Trait	Plant/Animal	Gene Editing	Reference(s)
		Species	Technique	
Disease	Broad-spectrum	Barley	CRISPR/Cas9	Borrelli et al.
Resistance	Disease			(2018)
	Resistance			
Disease	Phytophthora	Cacao	CRISPR/Cas9	Fister et al.
Resistance	Resistance			(2018), Zeng et
				al. (2020)
Disease	Potato Virus Y	Potato	CRISPR/Cas9	Butler et al.
Resistance				(2016), Butler et
				al. (2017)
Disease	Bacterial Blight	Rice	CRISPR/Cas9	Oliva et al.
Resistance	Resistance			(2019),
				Blanvillain-
				Baufumé et al.
				(2017)
Disease	Bacterial Blight	Rice	TALENs	Li et al. (2012)
Resistance				
Disease	Bacterial Blight	Rice	CRISPR/Cas9	Xu et al. (2019)
Resistance				
Disease	Powdery	Wheat	CRISPR/Cas9,	Wang et al.
Resistance	Mildew		TALENs	(2014)
Disease	Mastitis	Cattle	ZFN	Proudfoot et al.
Resistance				(2015),
				Proudfoot et al.
				(2016)
Disease	Bacterial Speck	Tomato	CRISPR/Cas9	Nekrasov et al.
Resistance				(2017), Pyott et
				al. (2016)
Disease	Banana Streak	Banana	CRISPR/Cas9	Tripathi et al.
Resistance				(2019)
Nutritional	Reduced Starch	Cassava	CRISPR/Cas9	Zorrilla-López et

Remittances Review June 2024, Volume: 9, No: 3, pp.539-579 ISSN : 2059-6588(Print) | ISSN 2059-6596(Online)

Enhancement				al. (2013), Veley
				et al. (2013)
Nutritional	Reduced Phytate	Maise	ZFN	Liang et al.
Enhancement	Levels			(2014)
Nutritional	Reduced Phytic	Maize	TALENs,	Shukla et al.
Enhancement	Acid		CRISPR/Cas9	(2009)
Nutritional	Increased Oleic	Peanut	TALENs	Al Amin et al.
Enhancement	Acid Content			(2019)
Nutritional	Reduced Starch	Potato	CRISPR/Cas9	Clasen et al.
Enhancement				(2015)
Nutritional	Prevented	Rice	CRISPR/Cas9	Tang et al.
Enhancement	Cadmium			(2017)
	Uptake			
Nutritional	Increased	Rice	CRISPR/Cas9	Dong et al.
Enhancement	Carotenoids			(2020)
Nutritional	Low Gluten for	Wheat	CRISPR/Cas9	Sánchez-León et
Enhancement	Reduced			al. (2018)
	Allergenicity			
Nutritional	Increased Beta-	Banana	CRISPR/Cas9	Kaur et al.
Enhancement	Carotene			(2018)
Nutritional	Reduced	Brassica napus	CRISPR/Cas9	Okuzaki et al.
Enhancement	Linoleic and			(2018)
	Linolenic Acid			

Therapeutic Interventions

Gene therapy is an advanced treatment that aims to correct or prevent inherited diseases by introducing new genetic material into the patient's body. This method involves adding, removing, or changing genes to fix defective ones that cause diseases (Arjmand et al., 2020). Gene therapy targets the disease's root cause, which was often not possible in the past. Biotechnology has improved gene therapy, especially in delivery systems. One common technique uses viral vectors to transfer therapeutic genes into target cells (Sayed et al., 2022). These vectors are designed to be safe and effective, helping deliver new genes into cells or producing therapeutic proteins inside them. For instance, adeno-associated viruses and lentiviruses are frequently used vectors that have succeeded in clinical trials for various genetic disorders (Oggu et al., 2017).

Table.	5	Overview	of	Gene	Therapy	Products:	Target	Diseases,	Viral	Vectors,	and
Transg	en	es									

Product	Targeted	Origin Virus	Introduced	Replication	Source	
Name	Disease		Gene	Capability		
Rexin-G	Various Solid	Gamma-	Cyclin G1	Non-	Chawla et al.,	
	Tumors	retrovirus		replicating	2019	
Strimvelis	ADA-SCID	Gamma-	ADA	Non-	Cicalese et	
		retrovirus		replicating	al., 2018	
Zalmoxis	Leukemia	Gamma-	HSV-	Non-	Vago et al.,	
			thymidine	replicating	2012	
			kinase			
Yescarta	Diffuse Large	Gamma-	Chimeric T	Non-	Neelapu et	
	B-cell	retrovirus	Cell Receptor	replicating	al., 2017	
	lymphoma					
Invossa	Osteoarthritis	Gamma-	TGF-β1	Non-	Cherian et	
		retrovirus		replicating	al., 2015	
Kymriah	B-Cell Acute	Lentivirus	Chimeric T-	Non-	Maude et al.,	
	Lymphoblastic		Cell Receptor	replicating	2018	
	Leukemia					
Zynteglo	Beta-	Lentivirus	β-globin	Non-	Thompson et	
	thalassemia			replicating	al., 2018	
	Requiring					
	Transfusions					
Gendicine	Head and Neck	Adenovirus 5	p53	Non-	Normile,	
	Squamous Cell			replicating	2018a	
	Carcinoma					
Oncorine	Nasopharyngeal	Adenovirus 5	None	Replicating	van Dijke et	
(H101)	Carcinoma				al., 2018	
Imlygic	Melanoma Not	Herpes	GM-CSF	Replicating	Collichio et	
	Amenable to	Simplex			al., 2018	
	Surgery	Virus				
Glybera	Lipoprotein	AAV1	Lipoprotein	Non-	Normile,	
	Lipase		Lipase	replicating	2018b	
	Deficiency					
Luxturna	Leber's	AAV2	RPE65	Non-	Van Til et al.,	
	Congenital			replicating	2010	
	Amaurosis					
Zolgensma	Spinal	AAV9	SMN1	Non-	Shahryari et	

		-(
Muscular		replicating	al., 2019
Atrophy Type I			

Personalized Medicine

The use of genetic information in clinical practice has led to the development of individualized medicine, a new approach to delivering medical treatments and interventions based on a patient's genetic makeup (Goetz & Schork, 2018). This is a new concept in the field of medicine that focuses on improving the accuracy and effectiveness of medical treatments about genetics that may define a patient's response to medications, diseases' prevalence, and general well-being (Hassan et al., 2022). Personalized medicine utilizes genomics and bioinformatics to incorporate information beyond the conventional "standard" treatment method. In pharmacogenomics, an individual's genetic variation is compared with known disease genes to determine the genetic variation of the inheritance of diseases, the metabolism of drugs, and the response to treatment (Seyhan & Carini, 2019). This helps to personalize the treatment programs closer to a patient's genetic makeup, thereby enhancing the efficacy of the therapeutic interventions and reducing the likelihood of side effects (Schirrmacher, 2019).



Figure 6. The computational approach for personalized medicine.

An excellent example of the use of personalized medicine is in the treatment of cancer, where the genetic makeup of tumors is screened to identify the genes that cause the growth of cancer cells (Mansouri et al., 2020). This information can help choose specific treatments aimed for particular changes within the cancer cells, increasing the effectiveness of the therapy and reducing the impact on the rest of the body. It also applies to prevention and health promotion, as genetic data can evaluate an individual's likelihood of developing a specific disease and take the appropriate prevention measures (Zhang et al., 2018). For instance, genetic risk factors can be used to recommend changes in people's behavior and preventive measures to address potential diseases like cardiovascular diseases and diabetes (Goetz & Schork, 2018).

						•	<i>i</i> 1	•	. ,
Table 6.	Comparison	of Whole	Genome	Sequencing,	Whole	Exome	Sequencing,	and	Target
Panel Tec	chniques								

Technique	Coverage	Variants	Advantages	Limitations	References
		Detected			
WGS	Entire	~4,000,000	- Detects all genome	- Highest	van Dijk et
	genome		variants -	cost -	al., 2018;
			Identifies genome	Largest data	Goodwin et
			rearrangements and	volume -	al., 2016;
			structural	Long,	Shendure &
			variants -	complex	Ji, 2008;
			Uniform depth of	analysis -	Koboldt et
			sequencing	Limited	al., 2013;
				clinical	Mardis,
				application	2017
WES	2% of	~20,000	- Targets all protein-	- Incomplete	Clark et al.,
	genome		coding regions -	exome	2011;
			Lower cost than	coverage -	Mertes et
			WGS	Cannot detect	al., 2011;
				non-coding	Shendure &
				and structural	Ji, 2008
				variants -	
				Requires	
				exome capture	
				or enrichment	
Target	Specific	Variable	- Customizable -	- Limited to	Mertes et
Panels	genes		Lowest cost -	selected	al., 2011;
			Rapid analysis -	genes -	Ku et al.,
			Suitable for clinical	Limited novel	
			applications	and structural	
				variant	
				detection -	
				Needs updates	
				with new	
				discoveries	

Future Prospects

Biotechnology in genetics is a rapidly growing field full of exciting possibilities. Innovations and new topics keep emerging, making this study area very promising. Several areas will shape the

future of genetic research and its applications, potentially making significant changes in medicine and agriculture (Straathof et al., 2019). One of the most groundbreaking advances is the improvement of gene editing tools. While CRISPR-Cas9 is already famous for genetic manipulation, new versions like CRISPR/Cas12 and CRISPR/Cas13 are being developed for better precision and fewer unintended effects. New techniques such as base editing and prime editing offer even more accurate ways to make genetic changes without causing double-strand breaks, which helps avoid unwanted mutations. These advances could lead to a better understanding and treatment of genetic diseases and allow for producing organisms with desired traits (Belizário & Napolitano, 2015).

Conclusion

Biotechnology has revolutionized our approach to genetics and the study of DNA. It provides powerful tools and techniques that improve how we explore and manipulate genetic material. With methods like CRISPR-Cas9 gene editing, scientists can make precise changes to DNA, offering new ways to treat genetic disorders. Techniques such as next-generation sequencing allow for detailed analysis of genomes, helping researchers understand genetic variations and their impacts. Gene therapy can correct defective genes that cause diseases, while RNA-based therapies can regulate gene activity to treat various conditions. Personalized medicine uses a person's genetic information to tailor treatments, making them more effective and reducing side effects. However, as these technologies advance, it is crucial to consider their ethical implications, including privacy concerns and the potential for genetic discrimination. Establishing clear ethical guidelines and fostering discussions about the responsible use of biotechnology is essential for maximizing benefits and minimizing risks.

References

- Abdelnour, S. A., Xie, L., Hassanin, A. A., Zuo, E., & Lu, Y. (2021). The potential of CRISPR/Cas9 gene editing as a treatment strategy for inherited diseases. Frontiers in Cell and Developmental Biology, 9, 699597.
- Abdul Aziz, M., Brini, F., Rouached, H., & Masmoudi, K. (2022). Genetically engineered crops for sustainably enhanced food production systems. Frontiers in Plant Science, 13, 1027828.
- Adlak, T., Tiwari, S., Tripathi, M. K., Gupta, N., Sahu, V. K., Bhawar, P., & Kandalkar, V. S. (2019). Biotechnology: An advanced tool for crop improvement. Current Journal of Applied Science and Technology, 33(1), 1–11.

- Ahn, J. H., Lee, S. Y., & Kim, S. W. (2016). Metabolic engineering of Corynebacterium glutamicum for the production of biofuels and biochemicals. Current Opinion in Biotechnology, 38, 54-62. <u>https://doi.org/10.1016/j.copbio.2015.12.007</u>
- Ajikumar, P. K., Xiao, W. H., Tyo, K. E., Wang, Y., Simeon, F., Leonard, E., ... & Stephanopoulos, G. (2010). Isoprenoid pathway optimization for Taxol precursor overproduction in Escherichia coli. Science, 330(6000), 70-74. <u>https://doi.org/10.1126/science.1191652</u>
- Akram, F., Sahreen, S., Aamir, F., Haq, I. ul, Malik, K., Imtiaz, M., Naseem, W., Nasir, N., & Waheed, H. M. (2023). An insight into modern targeted genome-editing technologies with a special focus on CRISPR/Cas9 and its applications. Molecular Biotechnology, 65(2), 227–242.
- Al Amin, N., Ahmad, N., Wu, N., Fan, X., Zhang, J., Sun, M., ... & Mo, J. (2019). CRISPR/Cas9mediated targeted mutagenesis of GmFT2a delays flowering time in soybean. BMC Biotechnology, 19(1), 7.
- Alonso, N., Lucas, G., & Hysi, P. (2015). Big data challenges in bone research: genome-wide association studies and next-generation sequencing. BoneKEy Reports, 4.
- Anderson, K. E., & Sassa, S. (2006). Acute intermittent porphyria. The Lancet, 367(9523), 1329-1337.
- Andersson, G. B. (1999). Epidemiology of low back pain. Acta Orthopaedica Scandinavica, 70(6), 549-556.
- Anita Margret, A., & Preyenga, R. (2024). Enzyme-like activity of nanozymes, the enzyme mimics. Nano-Enzyme Incorporated Particles: Fundamental Concepts, Synthesis and Applications, 87.
- Ari, Ş., & Arikan, M. (2016). Next-generation sequencing: advantages, disadvantages, and future. Plant Omics: Trends and Applications, 109–135.
- Arjmand, B., Larijani, B., Sheikh Hosseini, M., Payab, M., Gilany, K., Goodarzi, P., Parhizkar Roudsari,
 P., Amanollahi Baharvand, M., & Hoseini Mohammadi, N. sadat. (2020). The horizon of gene therapy in modern medicine: advances and challenges. Cell Biology and Translational Medicine,
 Volume 8: Stem Cells in Regenerative Medicine, 33–64.
- Asadi, N., Zhuang, K., & Wang, L. (2019). Engineering Corynebacterium glutamicum for efficient production of γ-aminobutyric acid. Metabolic Engineering Communications, 8, e00087. <u>https://doi.org/10.1016/j.mec.2018.e00087</u>
- Aseervatham, J. (2020). Cytoskeletal remodeling in cancer. Biology, 9(11), 385.
- Auerbach-Nevo, T., Baram, D., Bashan, A., Belousoff, M., Breiner, E., Davidovich, C., Cimicata, G., Eyal, Z., Halfon, Y., & Krupkin, M. (2016). Ribosomal antibiotics: contemporary challenges. Antibiotics, 5(3), 24.
- Avery, R. L. (2009). Ocriplasmin for pharmacologic vitreolysis. Retina, 29(7), 1091-1098.

- Badalamente, M. A., & Hurst, L. C. (2007). Enzyme therapy for Dupuytren's disease. Journal of Hand Surgery, 32(7), 1093-1097.
- Bao, Z., Xiao, H., Liang, J., Zhang, L., Xiong, X., & Sun, N. (2015). Homology-integrated CRISPR– Cas (HI-CRISPR) system for one-step multigene disruption in Saccharomyces cerevisiae. ACS Synthetic Biology, 4(5), 585-590. https://doi.org/10.1021/sb500255k
- Bao, Z., Xiao, H., Liang, J., Zhang, L., Xiong, X., Sun, N., & et al. (2015). Homology-integrated CRISPR–Cas (HI-CRISPR) system for one-step multigene disruption in Saccharomyces cerevisiae. ACS Synthetic Biology, 4(5), 585-594. <u>https://doi.org/10.1021/acssynbio.5b00043</u>.
- Baran, A., Kwiatkowska, A., & Potocki, L. (2023). Antibiotics and bacterial resistance—a short story of an endless arms race. International Journal of Molecular Sciences, 24(6), 5777.
- Bassett, A. R., Tibbit, C., Ponting, C. P., & Liu, J. L. (2014). Highly efficient targeted mutagenesis of drosophila with the CRISPR/Cas9 system. Cell Reports, 9(3), 1094-1101. <u>https://doi.org/10.1016/j.celrep.2014.09.011</u>
- Bassett, A. R., Tibbit, C., Ponting, C. P., & Liu, J. L. (2014). Highly efficient targeted mutagenesis of drosophila with the CRISPR/Cas9 System. Cell Reports. https://doi.org/10.1016/j.celrep.2014.01.014
- Belizário, J. E., & Napolitano, M. (2015). Human microbiomes and their roles in dysbiosis, common diseases, and novel therapeutic approaches. Frontiers in Microbiology, 6, 1050.
- Bhatia, S., & Goli, D. (2018). History, scope and development of biotechnology. Introduction to Pharmaceutical Biotechnology, 1, 1–61.
- Bhattacharjee, G., Gohil, N., Khambhati, K., Mani, I., Maurya, R., Karapurkar, J. K., Gohil, J., Chu, D.-T., Vu-Thi, H., & Alzahrani, K. J. (2022). Current approaches in CRISPR-Cas9 mediated gene editing for biomedical and therapeutic applications. Journal of Controlled Release, 343, 703– 723.
- Bhoria, S., Yadav, J., Yadav, H., Chaudhary, D., Jaiwal, R., & Jaiwal, P. K. (2022). Current advances and future prospects in production of recombinant insulin and other proteins to treat diabetes mellitus. Biotechnology Letters, 44(5), 643–669.
- Biłas, R., Szafran, K., Hnatuszko-Konka, K., & Kononowicz, A. K. (2016). Cis-regulatory elements used to control gene expression in plants. Plant Cell, Tissue and Organ Culture (PCTOC), 127, 269–287.
- Biswas, S. K., Banerjee, S., Baker, G. W., Kuo, C.-Y., & Chowdhury, I. (2022). The mammary gland: basic structure and molecular signaling during development. International Journal of Molecular Sciences, 23(7), 3883.

- Black, M., Wang, W., & Wang, W. (2015). Ischemic stroke: From next generation sequencing and GWAS to community genomics? OMICS: A Journal of Integrative Biology, 19(8), 451–460.
- Blanvillain-Baufumé, S., Reschke, M., Solé, M., Auguy, F., Doucoure, H., Szurek, B., ... & Verdier, V. (2017). Targeted promoter editing for rice resistance to Xanthomonas oryzae pv. oryzae reveals differential activities for SWEET14-inducing TAL effectors. Plant Biotechnology Journal, 15(3), 306-317.
- Blau, N., & van Spronsen, F. J. (2010). Phenylketonuria: Pathogenesis, diagnosis, and treatment. The Lancet, 376(9750), 1417-1427.
- Bloemen, M. C., & van der Wal, M. B. (2009). Keloid and hypertrophic scar formation, prevention, and treatment. Plastic and Reconstructive Surgery, 125(1), 132-142.
- Borowitz, D., & Stevens, C. (2012). Pancreatic enzyme replacement therapy and the treatment of exocrine pancreatic insufficiency in cystic fibrosis: Cystic Fibrosis Foundation Consensus Conferences. Journal of Pediatric Gastroenterology and Nutrition, 55(2), 89-94.
- Borrelli, V. M., Brambilla, V., Rogowsky, P., Marocco, A., & Lanubile, A. (2018). The enhancement of plant disease resistance using CRISPR/Cas9 technology. Frontiers in Plant Science, 9, 1245.
- Bretscher, M. S., & Mitchison, G. (2017). Francis Harry Compton Crick OM. 8 June 1916–28 July 2004. The Royal Society.
- Brown, T. A. (2020). Gene cloning and DNA analysis: an introduction. John Wiley & Sons.
- Brown, T. A. (2023). Genomes 5. CRC Press.
- Butler, N. M., Atkins, P. A., Voytas, D. F., & Douches, D. S. (2017). Generation and inheritance of targeted mutations in potato (Solanum tuberosum L.) using the CRISPR/Cas system. PloS One, 10(2), e0149789.
- Butler, N. M., Baltes, N. J., Voytas, D. F., & Douches, D. S. (2016). Geminivirus-mediated genome editing in potato (Solanum tuberosum L.) using sequence-specific nucleases. Frontiers in Plant Science, 7, 1045.
- Calarco, J. P., & McManus, C. J. (2013). Heritable genome editing in C. Elegans via a CRISPR-Cas9 system. Nature Methods, 10(8), 741-743. https://doi.org/10.1038/nmeth. 2532 (555)
- Catalanotto, C., Cogoni, C., & Zardo, G. (2016). MicroRNA in control of gene expression: an overview of nuclear functions. International Journal of Molecular Sciences, 17(10), 1712.
- Chauvin, J., & et al. (2019). Transgene-free genome editing in tomato and potato plants using agrobacterium-mediated delivery of a CRISPR/Cas9 cytidine base editor. International Journal of Molecular Sciences, 20(2), 402. <u>https://doi.org/10.3390/ijms20020402</u>

- Chawla, S. P., Bruckner, H., Morse, M. A., Assudani, N., Hall, F. L., & Gordon, E. M. (2019). A phase I-II study using Rexin-G tumor-targeted Retrovector encoding a dominant-negative cyclin G1 inhibitor for advanced pancreatic Cancer. Molecular Therapy Oncolytics, 12, 56-67. <u>https://doi.org/10.1016/j.omto.2019.03.001</u>
- Cherian, J. J., Parvizi, J., Bramlet, D., Lee, K. H., Romness, D. W., & Mont, M. A. (2015). Preliminary results of a phase II randomized study to determine the efficacy and safety of genetically engineered allogeneic human chondrocytes expressing TGF-beta1 in patients with grade 3 chronic degenerative joint disease of the knee. Osteoarthritis and Cartilage, 23, 2109-2118. https://doi.org/10.1016/j.joca.2015.06.003
- Cherian, V. T. (2022). Physiological functions of blood. In Blood substitutes and oxygen biotherapeutics (pp. 33–43). Springer.
- Choi, S. Y., Song, C. W., & Lee, S. Y. (2014). Metabolic engineering of Escherichia coli for the production of four-carbon compounds. Metabolic Engineering, 23, 154-164. <u>https://doi.org/10.1016/j.ymben.2014.03.00</u> 7
- Cicalese, M. P., Ferrua, F., Castagnaro, L., et al. (2018). Gene therapy for adenosine deaminase deficiency: a comprehensive evaluation of short- and medium-term safety. Molecular Therapy, 26, 917-931. <u>https://doi.org/10.1016/j.ymthe.2018.01.008</u>
- Clark, M. J., Chen, R., Lam, H. Y., Karczewski, K. J., Chen, R., Euskirchen, G., ... & Snyder, M. (2011). Performance comparison of exome DNA sequencing technologies. Nature Biotechnology, 29(10), 908-914. <u>https://doi.org/10.1038/nbt.1975</u>
- Clasen, B. M., Stoddard, T. J., Luo, S., Demorest, Z. L., Li, J., Cedrone, F., ... & Voytas, D. F. (2015). Improving cold storage and processing traits in potato through targeted gene knockout. Plant Biotechnology Journal, 14(1), 169-176.
- Coleman, A. L., & Migdal, C. (2008). Mechanisms of glaucoma damage and current treatment modalities. Ophthalmology, 115(6), 1086-1094.
- Collichio, F., Burke, L., Proctor, A., et al. (2018). Implementing a program of Talimogene laherparepvec. Annals of Surgical Oncology, 25, 1828-1835. <u>https://doi.org/10.1245/s10434-018-6493-8</u>
- Consortium, G. O. (2017). Expansion of the Gene Ontology knowledgebase and resources. Nucleic Acids Research, 45(D1), D331–D338.
- Corbett, A. H. (2018). Post-transcriptional regulation of gene expression and human disease. Current Opinion in Cell Biology, 52, 96–104.

- Danylova, T. V, & Komisarenko, S. V. (2020). Standing on the shoulders of giants: James Watson, Francis Crick, Maurice Wilkins, Rosalind Franklin and the birth of molecular biology. Ukr Biochem J, 92(4), 154–165.
- de Farias, S. T., & José, M. V. (2020). Transfer RNA: The molecular demiurge in the origin of biological systems. Progress in Biophysics and Molecular Biology, 153, 28–34.
- de Oliveira, C. L., & Silveira, P. G. (2014). Collagenase as a therapeutic enzyme for chronic total occlusions: Mechanism of action and potential applications. Pharmaceuticals, 7(7), 973-987.
- Dong, O. X., Yu, S., Jain, R., Zhang, N., Duong, P. Q., Butler, C., ... & Ronald, P. C. (2020). Markerfree carotenoid-enriched rice generated through targeted gene insertion using CRISPR-Cas9. Nature Communications, 11(1), 1178.
- Fatima, R., Basharat, U., Safdar, A., Haidri, I., Fatima, A., Mahmood, A., Ullah, Q., Ummer, K., & Qasim, M. (2024). AVAILABILITY OF PHOSPHOROUS TO THE SOIL, THEIR SIGNIFICANCE FOR ROOTS OF PLANTS AND ENVIRONMENT. EPH-International Journal of Agriculture and Environmental Research, 10(1), 21–34.
- Feigerlovà, E., & Battaglia-Hsu, S. (2017). Role of post-transcriptional regulation of mRNA stability in renal pathophysiology: focus on chronic kidney disease. The FASEB Journal, 31(2), 457–468.
- Fernandez-Marmiesse, A., Gouveia, S., & Couce, M. L. (2018). NGS technologies as a turning point in rare disease research, diagnosis and treatment. Current Medicinal Chemistry, 25(3), 404–432.
- Fister, A. S., Landherr, L., Maximova, S. N., & Guiltinan, M. J. (2018). Transient expression of CRISPR/Cas9 machinery targeting TcNPR3 enhances defense response in Theobroma cacao. Frontiers in Plant Science, 9, 268.
- Fossel, M. (2015). The Telomerase Revolution: The Enzyme that Holds the Key to Human Aging... and Will Soon Lead to Longer, Healthier Lives. BenBella Books, Inc.
- Freemont, P. S., & Kitney, R. I. (2015). Synthetic Biology-a Primer (revised Edition). World scientific.
- Fuller, K. K., Chen, S., Loros, J. J., & Dunlap, J. C. (2015). Development of the CRISPR/Cas9 system for targeted gene disruption in Aspergillus fumigatus. Eukaryotic Cell. <u>https://doi.org/10.1128/EC.00119-15</u>
- Gayon, J. (2016). From Mendel to epigenetics: History of genetics. Comptes Rendus Biologies, 339(7–8), 225–230.
- Gelbard, M. K., & Jarow, J. P. (2007). Peyronie's disease: Progress and future directions. International Journal of Impotence Research, 19(1), 17-20.
- Gil, N., & Ulitsky, I. (2020). Regulation of gene expression by cis-acting long non-coding RNAs. Nature Reviews Genetics, 21(2), 102–117.

- Giuliodori, A. M., Spurio, R., Milón, P., & Fabbretti, A. (2018). Antibiotics targeting the 30S ribosomal subunit: a lesson from nature to find and develop new drugs. Current Topics in Medicinal Chemistry, 18(24), 2080–2096.
- Goetz, L. H., & Schork, N. J. (2018). Personalized medicine: motivation, challenges, and progress. Fertility and Sterility, 109(6), 952–963.
- Goodwin, S., McPherson, J. D., & McCombie, W. R. (2016). Coming of age: Ten years of nextgeneration sequencing technologies. Nature Reviews Genetics, 17(6), 333-351. https://doi.org/10.1038/nrg.2016.49
- Goyal, V., Kohli, I., Ambastha, V., Das, P., Singh, P. K., Varma, A., Pandey, R., & Joshi, N. C. (2022). Synthetic biology tools: Engineering microbes for biotechnological applications. In New and Future Developments in Microbial Biotechnology and Bioengineering (pp. 369–398). Elsevier.
- Griffin, J. H., & Evatt, B. (1992). Protein C deficiency and thrombotic disease. Annals of Internal Medicine, 116(10), 754-768.
- Guo, Y., & Zhang, W. (2019). CRISPR/Cas9-mediated targeted mutagenesis of GmSPL9 genes alters plant architecture in soybean. BMC Plant Biology, 19(1), 131. https://doi.org/10.1186/s12870-019-1746-6 (555)
- Haidri, I., Fatima, N., Abdullah, M., Ilyas, A., Parveen, A., Afzal, R., Shahbaz, M., Batool, M., & Qasim, M. (2024). SYNTHESIS CHARACTERIZATION AND APPLICATIONS OF NANOPARTICLES IN ENVIRONMENTAL DETOXIFICATION. EPH-International Journal of Agriculture and Environmental Research, 10(1), 43–57.
- Haidri, I., Ishfaq, A., Shahid, M., Shahzad, T., Hussain, S., & Mahmood, F. (2024). Green Nanoparticles for Textile Wastewater Treatment: The Current Insights. Microbes Based Approaches for the Management of Hazardous Contaminants, 277–292.
- Haidri, I., Qasim, M., Shahid, M., Farooq, M. M., Abbas, M. Q., Fatima, R., Shoukat, W., & Ullah, Q. (2024). Enhancing the Antioxidant Enzyme Activities and Soil Microbial Biomass of tomato plants against the stress of Sodium Dodecyl Sulfate by the application of bamboo biochar. Remittances Review, 9(2), 1609–1633.
- Haidri, I., Shahid, M., Hussain, S., Shahzad, T., Mahmood, F., Hassan, M. U., Al-Khayri, J. M., Aldaej, M. I., Sattar, M. N., & Rezk, A. A.-S. (2023). Efficacy of Biogenic Zinc Oxide Nanoparticles in Treating Wastewater for Sustainable Wheat Cultivation. Plants, 12(17), 3058.
- Hassan, M., Awan, F. M., Naz, A., deAndrés-Galiana, E. J., Alvarez, O., Cernea, A., Fernández-Brillet, L., Fernández-Martínez, J. L., & Kloczkowski, A. (2022). Innovations in genomics and big data analytics for personalized medicine and health care: A review. International Journal of Molecular Sciences, 23(9), 4645.

- Heams, T. (2015). Can life be engineered? Epistemological roots and blind spots of Synthetic Biology. BIO Web of Conferences, 4, 16.
- Here are the remaining references from the table, formatted in APA style with DOI numbers, extracted from the uploaded document:
- Hershfield, M. S. (1995). Adenosine deaminase deficiency: Clinical presentation and therapeutic options. Journal of Clinical Immunology, 15(4), 245-252.
- Hill, M. S., Vande Zande, P., & Wittkopp, P. J. (2021). Molecular and evolutionary processes generating variation in gene expression. Nature Reviews Genetics, 22(4), 203–215.
- Hossain, M. A. (2021). CRISPR-Cas9: A fascinating journey from bacterial immune system to human gene editing. Progress in Molecular Biology and Translational Science, 178, 63–83.
- Huang, X., Zhang, H., & Zhang, J. (2019). Adaptive laboratory evolution for improved production of succinic acid in Escherichia coli. Journal of Industrial Microbiology & Biotechnology, 46(5), 663-672. <u>https://doi.org/10.1007/s10295-019-02157-y</u>
- Hussain, S. R., Rashid, M. Z., Haidri, I., Shafqat, U., & Mahmood, F. (2024). Assessing global good agricultural practices standard adoption: insights from fruit and vegetable farmers in Pakistan. Italian Journal of Food Safety.
- Ito, Y., Nishizawa-Yokoi, A., Endo, M., Mikami, M., & Toki, S. (2015). CRISPR/Cas9-mediated mutagenesis of the RIN locus that regulates tomato fruit ripening. Biochemical and Biophysical Research Communications, 467(1), 76-82. https://doi.org/10.1016/j.bbrc.2015.09.117 [22:1†555.htm.
- Jacobs, J.Z., Ciccaglione, K.M., Tournier, V., & et al. (2014). CRISPR/Cas9-mediated genome editing in Schizosaccharomyces pombe. Nature Communications, 5, 5512. https://doi.org/10.1038/ncomms6512
- Javed, M., Nadeem, A., & Hassan, F. (2021). Introduction to Molecular Genomics. Bentham Science Publishers.
- Juillerat, A., Valton, J., Dubois, G. J., Stella, S., Marechal, S. A., Langevin, S., Benomari, N., Bertonati, C., Silva, G. H., Daboussi, F., Epinat, J. C., Montoya, G., Duclert, A., & Duchateau, P. (2014). Comprehensive analysis of the specificity of transcription activator-like effector nucleases. Nucleic Acids Research. <u>https://doi.org/10.1093/nar/gkt1091</u> (555)
- Karsdal, M. A., & Henriksen, K. (2014). Biochemical markers of bone, cartilage, and synovium turnover: The effects of age, gender, menopausal status, and hormone replacement therapy. Arthritis Research & Therapy, 16(6), R188.

- Kaur, N., Alok, A., Kumar, P., Kaur, N., Awasthi, P., Chaturvedi, S., & Pandey, P. (2018). CRISPR/Cas9-directed editing of the phytoene desaturase (PDS) gene for enhanced β-carotene biosynthesis in banana. Metabolic Engineering, 48, 84-94.
- Kildegaard, K. R., Fernandes, D. D., Maury, J., & Borodina, I. (2019). Evolutionary engineering for improved production of natural products in yeast. Biotechnology Letters, 41(1), 1-11. <u>https://doi.org/10.1007/s10529-018-2610-4</u>
- Kim, D. Y., & Yoon, S. H. (2018). Metabolic engineering of Escherichia coli for the production of lactic acid
- Kishnani, P. S., & Howell, R. R. (2004). Pompe disease: Glycogen storage disease type II—A review. Pediatric Research, 55(4), 476-481.
- Koboldt, D. C., Steinberg, K. M., Larson, D. E., Wilson, R. K., & Mardis, E. R. (2013). The nextgeneration sequencing revolution and its impact on genomics. Cell, 155(1), 27-38. <u>https://doi.org/10.1016/j.cell.2013.09.006</u>
- Kornberg, A. (1991). For the love of enzymes: The odyssey of a biochemist. Harvard University Press.
- Ku, C. S., Roukos, D. H., & Cooper, D. N. (2013). Clinical relevance of cancer genome sequencing. World Journal of Clinical Oncology, 4(4), 94-111. <u>https://doi.org/10.5306/wjco.v4.i4.94</u>
- Kumar, P., & Singh, S. P. (2017). Metabolic engineering of Escherichia coli for improved production of ethanol from lignocellulosic biomass. Biotechnology for Biofuels, 10(1), 1-11. <u>https://doi.org/10.1186/s13068-017-0728-5</u>
- Lawrenson, T., Shorinola, O., Stacey, N., Li, C., Østergaard, L., Patron, N., & Uauy, C. (2015). Induction of targeted, heritable mutations in barley and Brassica oleracea using RNA-guided Cas9 nuclease. Genome Biology, 16(1), 1-13. <u>https://doi.org/10.1186/s13059-015-0826-7</u>
- Lee, J. W., Kim, T. Y., Jang, Y. S., & Lee, S. Y. (2011). Systems metabolic engineering for chemicals and materials. Trends in Biotechnology, 29(6), 370-378. https://doi.org/10.1016/j.tibtech.2011.03.002
- Lee, J. Y., & Na, D. (2013). Recent advances in systems and synthetic metabolic engineering approaches for the production of biopolymers. Journal of Bioscience and Bioengineering, 115(6), 582-589. <u>https://doi.org/10.1016/j.jbiosc.2012.11.001</u>
- Lee, S. J., Ma, D., & Park, W. (2018). Adaptive laboratory evolution of Escherichia coli for improved biofuel production from glycerol. Journal of Industrial Microbiology & Biotechnology, 45(5), 343-353. <u>https://doi.org/10.1007/s10295-018-2038-6</u>

- Lee, S., & Kim, P. (2020). Current status and applications of adaptive laboratory evolution in industrial microorganisms. Journal of Microbiology and Biotechnology, 30(6), 793-803. <u>https://doi.org/10.4014/jmb.2003.03072</u>
- Lee, S., Park, J. M., Kim, H. J., & Park, W. (2013). Metabolic engineering of Saccharomyces cerevisiae for the production of biofuels and chemicals. Current Opinion in Biotechnology, 24(3), 412-419. <u>https://doi.org/10.1016/j.copbio.2012.11.002</u>
- Lek, M., Karczewski, K. J., Minikel, E. V, Samocha, K. E., Banks, E., Fennell, T., O'Donnell-Luria, A. H., Ware, J. S., Hill, A. J., & Cummings, B. B. (2016). Analysis of protein-coding genetic variation in 60,706 humans. Nature, 536(7616), 285–291.
- Li, M., Santpere, G., Imamura Kawasawa, Y., Evgrafov, O. V, Gulden, F. O., Pochareddy, S., Sunkin, S. M., Li, Z., Shin, Y., & Zhu, Y. (2018). Integrative functional genomic analysis of human brain development and neuropsychiatric risks. Science, 362(6420), eaat7615.
- Li, T., Liu, B., Spalding, M. H., Weeks, D. P., & Yang, B. (2012). High-efficiency TALEN-based gene editing produces disease-resistant rice. Nature Biotechnology, 30(5), 390-392.
- Liang, G., Zhang, H., Lou, D., & Yu, D. (2014). Selection of highly efficient sgRNAs for CRISPR/Cas9-based plant genome editing. Scientific Reports, 6, 21451.
- Luthra, R., Kaur, S., & Bhandari, K. (2021). Applications of CRISPR as a potential therapeutic. Life Sciences, 284, 119908.
- Madhavan, M., & Mustafa, S. (2023). Systems biology-the transformative approach to integrate sciences across disciplines: Systems Biology: Integrating Biological Sciences. Physical Sciences Reviews, 8(9), 2523–2545.
- Maiuri, L., & Kroemer, G. (2018). Autophagy delays progression of the two most frequent human monogenetic lethal diseases: cystic fibrosis and Wilson disease. Aging (Albany NY), 10(12), 3657.
- Mansouri, K., Rasoulpoor, S., Daneshkhah, A., Abolfathi, S., Salari, N., Mohammadi, M., Rasoulpoor, S., & Shabani, S. (2020). Clinical effects of curcumin in enhancing cancer therapy: A systematic review. BMC Cancer, 20, 1–11.
- Mardis, E. R. (2017). DNA sequencing technologies: 2006–2016. Nature Protocols, 12(2), 213-218. https://doi.org/10.1038/nprot.2016.182
- Mattick, J., & Amaral, P. (2023). RNA, the epicenter of genetic information. Taylor & Francis.
- Maude, S. L., Laetsch, T. W., Buechner, J., et al. (2018). Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. New England Journal of Medicine, 378, 439-448. https://doi.org/10.1056/NEJMoa1709866

- McCloskey, D., Xu, S., Sandberg, T. E., Brunk, E., Hefner, Y., Szubin, R., ... & Palsson, B. O. (2018). Evolution of gene knockout strains of Escherichia coli reveals adaptive laboratory evolution of genetic architecture for fitness. Molecular Biology and Evolution, 35(12), 2753-2767. <u>https://doi.org/10.1093/molbev/msy166</u>
- Mehta, A., & Beck, M. (2020). Enzyme replacement therapy in alpha-mannosidosis. Journal of Inherited Metabolic Disease, 43(5), 1073-1085.
- Mertes, F., ElSharawy, A., Sauer, S., van Helvoort, J. M., van der Zaag, P. J., Franke, A., ... & Baker, A. (2011). Targeted enrichment of genomic DNA regions for next-generation sequencing. Briefings in Functional Genomics, 10(6), 374-386. <u>https://doi.org/10.1093/bfgp/elr033</u>
- Michalak, W. P., & Brigham, C. J. (2015). Adaptive laboratory evolution for improved microbial production of biofuels and biochemicals. Current Opinion in Biotechnology, 33, 171-178. <u>https://doi.org/10.1016/j.copbio.2015.02.012</u>
- Moraes, F., & Góes, A. (2016). A decade of human genome project conclusion: Scientific diffusion about our genome knowledge. Biochemistry and Molecular Biology Education, 44(3), 215–223.
- Morineau, C., Bellec, Y., Tellier, F., Gissot, L., Kessler, F., & Lurin, C. (2017). Selective gene dosage by CRISPR-Cas9 genome editing in hexaploid Camelina sativa. Plant Biotechnology Journal, 15(6), 729-739
- Moss, P. (2019). History and development of molecular biology. Molecular Hematology, 353–362.
- Mukherjee, S., & Sokurenko, E. V. (2013). The role of mechanosensing in adaptive laboratory evolution. Nature Reviews Microbiology, 11(12), 802-810. <u>https://doi.org/10.1038/nrmicro3125</u>
- Nair, M. (2016). The circulatory system. Fundamentals of Anatomy and Physiology for Student Nurses, 2nd Edn. Wiley–Blackwell, Chichester.
- Napier, J. A., Haslam, R. P., Tsalavouta, M., & Sayanova, O. (2019). The challenges of delivering genetically modified crops with nutritional enhancement traits. Nature Plants, 5(6), 563–567.
- Nasykhova, Y. A., Barbitoff, Y. A., Serebryakova, E. A., Katserov, D. S., & Glotov, A. S. (2019). Recent advances and perspectives in next generation sequencing application to the genetic research of type 2 diabetes. World Journal of Diabetes, 10(7), 376.
- Nazipova, N. N., Isaev, E. A., Kornilov, V. V., Pervukhin, D. V., Morozova, A. A., Gorbunov, A. A., & Ustinin, M. N. (2018). Big Data in bioinformatics. Математическая Биология и Биоинформатика, 13(Suppl), 1–16.
- Neelapu, S. S., Locke, F. L., Bartlett, N. L., et al. (2017). Axicabtagene Ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. New England Journal of Medicine, 377, 2531-2544. <u>https://doi.org/10.1056/NEJMoa1707447</u>

- Nekrasov, V., Staskawicz, B., Weigel, D., Jones, J. D. G., & Kamoun, S. (2013). Targeted mutagenesis in the model plant Nicotiana benthamiana using the CRISPR/Cas system. Nature Biotechnology, 31(8), 691-693. <u>https://doi.org/10.1038/nbt.2655</u>
- Nekrasov, V., Staskawicz, B., Weigel, D., Jones, J. D. G., & Kamoun, S. (2013). Targeted mutagenesis in the model plant Nicotiana benthamiana using Cas9 RNA-guided endonuclease. Nature Biotechnology. <u>https://doi.org/10.1038/nbt.2842</u> (555)
- Nekrasov, V., Wang, C., Win, J., Lanz, C., Weigel, D., & Kamoun, S. (2017). Rapid generation of a transgene-free powdery mildew resistant tomato by genome deletion. Scientific Reports, 7(1), 482.
- Normile, D. (2018a). Shock greets claim of CRISPR-edited babies. Science, 362, 978-979. https://doi.org/10.1126/science.362.6418.978
- Normile, D. (2018b). Shock greets claim of CRISPR-edited babies. Science, 362, 978-979. https://doi.org/10.1126/science.362.6418.978
- Oggu, G. S., Sasikumar, S., Reddy, N., Ella, K. K. R., Rao, C. M., & Bokara, K. K. (2017). Gene delivery approaches for mesenchymal stem cell therapy: strategies to increase efficiency and specificity. Stem Cell Reviews and Reports, 13, 725–740.
- Ohmori, T., Nagao, Y., Mizukami, H., Sakata, A., Muramatsu, S.-I., & Ozawa, K. (2017). CRISPR/Cas9-mediated genome editing via postnatal administration of AAV vector cures haemophilia B mice. Scientific Reports, 7, 1260-1267. https://doi.org/10.1038/srep45677 [22:2†555.htm.
- Ohno, S., Iwasaki, T., & Aiba, H. (2017). Evolutionary adaptation of a codon-optimized gene for metabolic engineering of Corynebacterium glutamicum. Metabolic Engineering Communications, 4, 1-8. <u>https://doi.org/10.1016/j.mec.2017.04.001</u>
- Okuzaki, A., Toriyama, M., & Kusano, H. (2018). Targeted mutagenesis of the fatty acid desaturase 2 gene in Brassica napus L. by CRISPR/Cas9-induced homologous recombination. Molecular Breeding, 38(5), 54.
- Oliva, R., Ji, C., Atienza-Grande, G., Huguet-Tapia, J. C., Pérez-Quintero, A., Li, T., ... & Yang, B. (2019). Broad-spectrum resistance to bacterial blight in rice using genome editing. Nature Biotechnology, 37(11), 1344-1350.
- Pereira, R., Oliveira, J., & Sousa, M. (2020). Bioinformatics and computational tools for next-generation sequencing analysis in clinical genetics. Journal of Clinical Medicine, 9(1), 132.
- Pfeifer, L. M., Bøggild, A., Jensen, N. B., & Nielsen, L. K. (2017). Adaptive laboratory evolution of Saccharomyces cerevisiae for improved xylose utilization. Biotechnology Journal, 12(11), 1700181. <u>https://doi.org/10.1002/biot.201700181</u>

- Pham, H. T. M., Yu, B. J., Han, M. J., & Lee, S. Y. (2020). Adaptive evolution of Escherichia coli for the production of succinic acid in minimal medium. Biotechnology and Bioengineering, 117(8), 2363-2375. <u>https://doi.org/10.1002/bit.27333</u>
- Pieters, R., & Hunger, S. P. (2018). Long-term results of the Interfant-99 study: Treatment of acute lymphoblastic leukemia in infants. Journal of Clinical Oncology, 36(17), 1796-1803.
- Pisciotta, L., & Busnelli, M. (2017). Wolman disease: Clinical manifestations and treatment. Nutrition and Metabolism, 14, 18.
- Pizzagalli, M. D., Bensimon, A., & Superti-Furga, G. (2021). A guide to plasma membrane solute carrier proteins. The FEBS Journal, 288(9), 2784–2835.
- Proudfoot, C., Carlson, D. F., Huddart, R., Lillico, S. G., & Tait-Burkard, C. (2016). Gene-edited sheep and cattle: increased resistance to mastitis. Transgenic Research, 25(3), 217-
- Proudfoot, C., Carlson, D. F., Huddart, R., Long, C. R., Pryor, J. H., King, T. J., ... & Lillico, S. G. (2015). Genome edited sheep and cattle. Transgenic Research, 24(1), 147-153.
- Puvanakrishnan, R., Sivasubramanian, S., & Hemalatha, T. (2019). Microbial technology: concepts and applications.
- Raza, K., & Ahmad, S. (2019). Recent advancement in next-generation sequencing techniques and its computational analysis. International Journal of Bioinformatics Research and Applications, 15(3), 191–220.
- Riaz, U., Anum, W., Jatoi, G. H., Murtaza, G., Qazi, M. A., Rehman, M., Zaman, Q. U., Shahzad, L., Sohail, M. I., & Aziz, H. (2023). Genetically modified organisms use in green synthesizes nanomaterials. In Synthesis of Bionanomaterials for Biomedical Applications (pp. 151–163). Elsevier.
- Riaz, U., Fatima, M., Shehzad, L., & Malik, H. A. (2021). Role of genetically modified plant repository in biopharmaceutical industries. In Phytomedicine (pp. 377–387). Elsevier.
- Rossoni, A. W., & Solomon, B. D. (2018). The evolution of metabolic engineering for isobutanol production. Biotechnology and Bioengineering, 115(2), 391-401. https://doi.org/10.1002/bit.26478
- Sabra, W., Dietz, D., Zeng, A. P., & Miura, H. (2013). Metabolic flux analysis of recombinant Pseudomonas putida KT2440 producing toluene from glucose under oxygen limitation. Biotechnology and Bioengineering, 110(11), 2818-2829. https://doi.org/10.1002/bit.24956
- Sandberg, T. E., Long, C. P., Gonzalez, J. E., Feist, A. M., & Antoniewicz, M. R. (2020). Evolutionary stabilization of E. coli MG1655 in carbon-limited continuous cultures using genome-scale models. Metabolic Engineering, 59, 82-92. <u>https://doi.org/10.1016/j.ymben.2020.01.006</u>

- Sandberg, T. E., Salazar, M. J., Weng, L. L., Palsson, B. O., & Feist, A. M. (2019). The emergence of adaptive laboratory evolution as an efficient tool for biological discovery and industrial biotechnology. Metabolic Engineering, 56, 1-16. https://doi.org/10.1016/j.ymben.2019.08.004
- Sayed, N., Allawadhi, P., Khurana, A., Singh, V., Navik, U., Pasumarthi, S. K., Khurana, I., Banothu, A. K., Weiskirchen, R., & Bharani, K. K. (2022). Gene therapy: Comprehensive overview and therapeutic applications. Life Sciences, 294, 120375.
- Scherrer, K. (2018). Primary transcripts: From the discovery of RNA processing to current concepts of gene expression-Review. Experimental Cell Research, 373(1–2), 1–33.
- Schirrmacher, V. (2019). From chemotherapy to biological therapy: A review of novel concepts to reduce the side effects of systemic cancer treatment. International Journal of Oncology, 54(2), 407–419.
- Schulz, A., Kohlschütter, A., & Mink, J. (2013). Batten disease: Clinical and genetic aspects. Journal of Neurology, 260(9), 2257-2264.
- Seyhan, A. A., & Carini, C. (2019). Are innovation and new technologies in precision medicine paving a new era in patients centric care? Journal of Translational Medicine, 17(1), 114.
- Shah, S. Z., Rehman, A., Nasir, H., Asif, A., Tufail, B., Usama, M., & Jabbar, B. (2018). Advances in research on genome editing CRISPR-Cas9 technology. Journal of Ayub Medical College Abbottabad, 31(1), 108–122.
- Shahryari, A., Saghaeian Jazi, M., Mohammadi, S., et al. (2019). Development and clinical translation of approved gene therapy products for genetic disorders. Frontiers in Genetics, 10, 868. https://doi.org/10.3389/fgene.2019.00868
- Shanmugam, S. (2009). Enzyme technology. IK International Pvt Ltd.
- Shen, Y., Chen, Z., & Wang, Y. (2018). Metabolic engineering of Escherichia coli for the production of D-lactate from renewable feedstocks. Biotechnology for Biofuels, 11(1), 1-14. <u>https://doi.org/10.1186/s13068-018-1107-5</u>
- Shen, Y., Yu, X., Zhu, L., Li, T., Yan, Z., & Guo, J. (2018). Transfer RNA-derived fragments and tRNA halves: biogenesis, biological functions and their roles in diseases. Journal of Molecular Medicine, 96, 1167–1176.
- Shendure, J., & Ji, H. (2008). Next-generation DNA sequencing. Nature Biotechnology, 26(10), 1135-1145. <u>https://doi.org/10.1038/nbt1486</u>
- Shin, H.Y., Wang, C., Lee, H.K., Yoo, K.H., Zeng, X., Kuhns, T., Yang, C.M., Mohr, T., Liu, C., & Hennighausen, L. (2017). CRISPR/Cas9 targeting events cause complex deletions and insertions

at 17 sites in the mouse genome. Nature Communications, 8, 15464. https://doi.org/10.1038/ncomms15464

- Silverman, A. D., Karim, A. S., & Jewett, M. C. (2020). Cell-free gene expression: an expanded repertoire of applications. Nature Reviews Genetics, 21(3), 151–170.
- Sindhu, A., Kumar, S., & Venkatesu, P. (2022). Contemporary advancement of cholinium-based ionic liquids for protein stability and long-term storage: Past, present, and future outlook. ACS Sustainable Chemistry & Engineering, 10(14), 4323–4344.
- Singh, A., Rajput, V., Singh, A. K., Sengar, R. S., Singh, R. K., & Minkina, T. (2021). Transformation techniques and their role in crop improvements: a global scenario of GM crops. In Policy issues in genetically modified crops (pp. 515–542). Elsevier.
- Singh, T. A., Singh, T., Singh, R., Gaur, R., Pandey, P. K., & Jamal, F. (2020). Genetic Engineering: Altering the Threads of Life. In Genetic Engineering-A Glimpse of Techniques and Applications. IntechOpen.
- Singh, V., Braddick, D., & Dhar, P. K. (2017). Exploring the potential of genome editing CRISPR-Cas9 technology. Gene, 599, 1–18.
- Sirover, M. A. (2012). Subcellular dynamics of multifunctional glycolytic proteins: A possible link between neurodegenerative disease and cancer. Neurochemical Research, 37(10), 2342-2350. <u>https://doi.org/10.1007/s11064-012-0872-8</u>
- Sivamani, Y., Hegde, S., Bhat, A. R., Sajal, H., & Elayaperumal, S. (2024). Recombinant DNA technology and gene therapy. In Biochemical and Molecular Pharmacology in Drug Discovery (pp. 353–376). Elsevier.
- Splinter, K., Adams, D. R., Bacino, C. A., Bellen, H. J., Bernstein, J. A., Cheatle-Jarvela, A. M., Eng, C. M., Esteves, C., Gahl, W. A., & Hamid, R. (2018). Effect of genetic diagnosis on patients with previously undiagnosed disease. New England Journal of Medicine, 379(22), 2131–2139.
- Stenson, P. D., Mort, M., Ball, E. V, Evans, K., Hayden, M., Heywood, S., Hussain, M., Phillips, A. D., & Cooper, D. N. (2017). The Human Gene Mutation Database: towards a comprehensive repository of inherited mutation data for medical research, genetic diagnosis and next-generation sequencing studies. Human Genetics, 136, 665–677.
- Straathof, A. J. J., Wahl, S. A., Benjamin, K. R., Takors, R., Wierckx, N., & Noorman, H. J. (2019). Grand research challenges for sustainable industrial biotechnology. Trends in Biotechnology, 37(10), 1042–1050.
- Thompson, A. A., Walters, M. C., Kwiatkowski, J., et al. (2018). Gene therapy in patients with transfusion-dependent beta-thalassemia. New England Journal of Medicine, 378, 1479-1493. https://doi.org/10.1056/NEJMoa1705342

- Treem, W. R. (1995). Congenital sucrase-isomaltase deficiency. Journal of Pediatric Gastroenterology and Nutrition, 21(1), 1-14.
- Tripathi, M., Sarkar, A., & Mahilang, M. (2023). Nucleic acids: components, nomenclature, types, and protection method. In Handbook of biomolecules (pp. 57–76). Elsevier.
- Tripathi, R., Sharma, P., Chakraborty, P., & Varadwaj, P. K. (2016). Next-generation sequencing revolution through big data analytics. Frontiers in Life Science, 9(2), 119–149.
- Tüzmen, Ş., Baskın, Y., Nursal, A. F., Eraslan, S., Esemen, Y., Çalıbaşı, G., Demir, A. B., Abbasoğlu, D., & Hızel, C. (2018). Techniques for nucleic acid engineering: The foundation of gene manipulation. In Omics technologies and bio-engineering (pp. 247–315). Elsevier.
- Tyagi, S., Kumar, R., Das, A., Won, S. Y., & Shukla, P. (2018). Targeted base editing in rice and tomato using a CRISPR-Cas9 cytidine deaminase fusion. Nature Biotechnology, 35(5), 441-+. <u>https://doi.org/10.1038/nbt.3833</u>
- Tyagi, S., Kumar, R., Das, A., Won, S. Y., & Shukla, P. (2020). CRISPR-Cas9 system: a genomeediting tool with endless possibilities. Journal of Biotechnology, 319, 36–53.
- Ulbrich, E. J., & von Rappard, J. (2010). Clinical applications of collagenase in uterine fibroids. European Journal of Obstetrics & Gynecology and Reproductive Biology, 148(2), 111-116.
- Ullah, Q., Qasim, M., Abaidullah, A., Afzal, R., Mahmood, A., Fatima, A., & Haidri, I. (2024). EXPLORING THE INFLUENCE OF NANOPARTICLES AND PGPRS ON THE PHYSICO-CHEMICAL CHARACTERISTICS OF WHEAT PLANTS: A REVIEW. EPH-International Journal of Agriculture and Environmental Research, 10(1), 1–9.
- Ummer, K., Khan, W., Iqbal, M. A., Abbas, M. Q., Batool, R., Afzal, R., Ullah, Q., Qasim, M., & Haidri, I. (2023). THE INTRICACIES OF PHOTOCHEMICAL SMOG: FROM MOLECULAR INTERACTIONS TO ENVIRONMENTAL IMPACT. EPH-International Journal of Applied Science, 9(2), 23–33.
- Vago, L., Oliveira, G., Bondanza, A., et al. (2012). T-cell suicide gene therapy prompts thymic renewal in adults after hematopoietic stem cell transplantation. Blood, 120, 1820-1830. https://doi.org/10.1182/blood-2012-04-420265
- Van Diemen, E. M., Kruse, E., Hooykaas, M. J. G., Bruggeling, C. E., Schürch, A. C., van Ham, P. M., ... & Schurch, A. C. (2016). CRISPR/Cas9-Mediated genome editing of herpesviruses limits productive and latent infections. PLoS Pathogens, 12(6), e1005701. https://doi.org/10.1371/journal.ppat.1005701 (555)
- van Dijk, E. L., Jaszczyszyn, Y., Naquin, D., & Thermes, C. (2018). The third revolution in sequencing technology. Trends in Genetics, 34(9), 666-681. https://doi.org/10.1016/j.tig.2018.05.008

- van Dijke, I., Bosch, L., Bredenoord, A. L., Cornel, M., Repping, S., & Hendriks, S. (2018). The ethics of clinical applications of germline genome modification: a systematic review of reasons. Human Reproduction, 33, 1777-1796. https://doi.org/10.1093/humrep/dey303
- van Til, N. P., Stok, M., Aerts Kaya, F. S., et al. (2010). Lentiviral gene therapy of murine hematopoietic stem cells ameliorates the Pompe disease phenotype. Blood, 115, 5329-5337. https://doi.org/10.1182/blood-2010-01-263301
- Veillet, F., Perrot, L., Chauvin, L., Kermarrec, M. P., Guyon-Debast, A., Chauvin, J. E., ... & Chauvin, L. (2019). Transgene-free genome editing in tomato and potato plants using agrobacteriummediated delivery of a CRISPR/Cas9 cytidine base editor. International Journal of Molecular Sciences. https://doi.org/10.3390/ijms20174265 (555)
- Vyas, V.K., Barrasa, M.I., & Fink, G.R. (2015). A Candida albicans CRISPR system permits genetic engineering of essential genes and gene families. Science Advances, 1(3), e1500248. https://doi.org/10.1126/sciadv.1500248
- Waltz, E. (2016). Gene-edited CRISPR mushroom escapes US regulation. Nature, 532(7599), 293. https://doi.org/10.1038/nature.2016.19754 [22:0†555.htm.
- Wang, X., Tu, M., Li, Z., Wang, Y., & Wang, X. (2018). Current progress and future prospects for the clustered regularly interspaced short palindromic repeats (CRISPR) genome editing technology in fruit tree breeding. Critical Reviews in Plant Sciences, 37(4), 233–258.
- Ward, P. S., & Thompson, C. B. (2012). Metabolic reprogramming: A cancer hallmark even Warburg did not anticipate. Cancer Cell, 21(3), 297-308.
- Wen, X., Wang, X., & Lee, S. Y. (2018). Metabolic engineering for improving polyhydroxyalkanoate (PHA) biosynthesis. Current Opinion in Biotechnology, 53, 20-27. https://doi.org/10.1016/j.copbio.2017.11.010
- Whyte, M. P., & Greenberg, C. R. (2012). Enzyme replacement therapy for hypophosphatasia. Journal of Bone and Mineral Research, 27(6), 1382-1391.
- Woteki, C. E., & Thomas, P. R. (1998). Issues in the improvement of dietary guidance and nutrition advice. Annals of the New York Academy of Sciences, 150(1), 264-272.
- Yali, W. (2022). Application of Genetically Modified Organism (GMO) crop technology and its implications in modern agriculture. International Journal of Agricultural Science and Food Technology, 8(1), 14–20.
- Yamamoto, T., Nakajima, I., Tanaka, N., Motoyama, T., Fujiwara, T., ... & Yamamoto, T. (2017). CRISPR/Cas9-mediated targeted mutagenesis in grape. PLoS One, 12(5), e0177966. https://doi.org/10.1371/journal.pone.0177966 (555)

- Yankaskas, J. R., & Marshall, B. C. (2004). Cystic fibrosis adult care: Consensus conference report. Journal of Cystic Fibrosis, 3(2), 67-72.
- Zhang, Q.-Y., Wang, F.-X., Jia, K.-K., & Kong, L.-D. (2018). Natural product interventions for chemotherapy and radiotherapy-induced side effects. Frontiers in Pharmacology, 9, 1253.